

Effect of Lead stress on morphology and chlorophyll content of chickpea seedlings

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Abstract

Heavy metal contamination in soil, particularly lead (Pb), is a growing environmental issue worldwide including India. Pb, a toxic metal with no biological function, is known to adversely affect both plant and animal life by disrupting cellular functions and causing organ damage. Pb toxicity in plants results in reduced growth, metabolic disturbances and compromised photosynthetic activity.

In this study, morphological, biochemical and physiological changes in chickpea seedlings have been studied under varying concentrations of Pb. Chickpea seedlings showed a negative response to Pb(NO₃)₂ on morphological parameters as well as biochemical components. This indicates that lead is a toxic element for plant growth and development. Lead had a substantial impact on the chlorophyll a, b and total chlorophyll content.

Keywords: Abiotic stress, heavy metal stress, lead nitrate [Pb(NO₃)₂], chlorophyll, fresh weight, dry weight.

Introduction

Heavy metal contamination in soil, particularly lead (Pb), is a growing environmental issue worldwide including India. Pb, a toxic metal with no biological function, accumulates in the environment due to industrialization and anthropogenic activities. It is known to adversely affect both plant and animal life by disrupting cellular functions and causing organ damage. Pb toxicity in plants results in reduced growth, metabolic disturbances and compromised photosynthetic activity. Heavy metals in biological systems have been found to affect cellular organelles and components including cell membranes, mitochondria, lysosomes, endoplasmic reticulum and nuclei, as well as some enzymes involved in metabolism, detoxification and damage repair⁶.

DNA damage and conformational changes leading to cell cycle modulation, carcinogenesis, or apoptosis have been caused due to metal ions interacting with cellular components such as DNA and nuclear proteins¹. In the lignin cellulose biomass samples (Mbwazirume and Nakyinyika peels) acquired from bananas, the presence of lead was reported⁷. Lichens were used as bioindicators to determine the different isotopes of Pb since decades³. Pb-affected soils contain 400–800 mg kg⁻¹ of lead whereas in industrialized areas, it may extend up to 1000 mg kg⁻¹ of soil. The negative

effects of Pb have been described on the growth and expansion of plants⁴.

Material and Methods

Seed Germination: Chickpea seeds were grown hydroponically in nutrient solution containing varying concentrations of lead nitrate Pb(NO₃)₂ ranging from 0 to 100 μM in a plant growth chamber at 28°C and 80% relative humidity for 10 days to assess the effects on root, stem and leaf growth.

Lead treatment: Seedlings were treated with different concentrations of Pb(NO₃)₂. They were grown in separate dishes filled with nutrient solution DW, 0 μM, 25 μM, 50 μM, 75 μM and 100 μM of Pb(NO₃)₂.

Measurement of length: Seedlings were harvested on the 10th day for measurement of length. Leaves, roots and shoots were collected separately. Their length was then measured in centimeters with the help of a scale.

Determination of fresh weight: 10th-day-old seedlings were harvested for determination of fresh weight. Seedlings were blotted dry and their fresh weights were measured.

Determination of dry weight: After determining the fresh weight, these seedlings were incubated for 72 hours at 65 degrees centigrade and weights were measured.

Extraction of chlorophyll: One gram of fresh leaves was collected and ground with 20–40 ml of 80% acetone. The mixture was centrifuged at 5000–10000 rpm for 5 mins. This procedure was repeated with the supernatant till the residue becomes colorless. The absorbance of the solution was recorded at 645 nm and 663 nm against the solvent (acetone) blank.

Estimation of Chlorophyll Content: The concentrations of chlorophyll a, chlorophyll b and total chlorophyll were calculated using the following equation:

$$\text{Total Chlorophyll: } 20.2(A645) + 8.02(A663)$$

$$\text{Chlorophyll a: } 12.7(A663) - 2.69(A645)$$

$$\text{Chlorophyll b: } 22.9(A645) - 4.68(A663)$$

Results and Discussion

Morphological studies: Measurements of roots, stems and leaves of the chickpea seedlings were taken at different concentrations of Pb(NO₃)₂ on the 10th day which are presented in table 1 and fig. 1. The observation was that the length of the leaf, root and stem of gram seedlings decreased

with increased concentration of $\text{Pb}(\text{NO}_3)_2$ compared to control plants. There is about 13.7%, 34.4% and 22.8% decrease in root length at 25 μm , 50 μm and 100 μm respectively. Stem length was decreased by about 12.34% and 20.14% at 25 μm and 100 μm $\text{Pb}(\text{NO}_3)_2$ respectively. There is also a significant decrease in leaf length with a significant increase in the concentration of $\text{Pb}(\text{NO}_3)_2$. This result is similar to the earlier findings of Jaishankar et al⁴.

Effect of lead on fresh weight and dry weight: Table 2 and fig. 2 demonstrate the dry weight and fresh weight in chickpea seedlings. It was observed that there is a decline in fresh weight at higher concentrations of lead, while dry weight increased significantly as compared to the control. It is in accordance with similar reports of other heavy metal exposure also.

Effect of lead on chlorophyll pigment: Total chlorophyll was found to be decreasing as the concentration of $\text{Pb}(\text{NO}_3)_2$ increases in the samples. Treatment at 25 μm and 100 μm

recorded a significant decline of 49.53% and 49.62% respectively. In chlorophyll. Chlorophyll a and b also declined at 100 μm conc. of $\text{Pb}(\text{NO}_3)_2$ in comparison to the control. The results were in accordance with the previous studies. Chlorophyll b is more adversely affected as compared to chlorophyll a by Pb toxicity⁵. Contamination of plants with lead increased the activity of chlorophyllase, which led to chlorophyll breakdown².

Conclusion

The present result allows us to conclude that chickpea seedlings showed a negative response to $\text{Pb}(\text{NO}_3)_2$ on morphological parameters as well as biochemical components. This indicates that lead is a toxic element for plant growth and development. Lead had a substantial impact on the chlorophyll. Uptake of lead is not only toxic, but it also affects the uptake of other essential elements which may cause a deficit of other nutrients in plants.

Table 1
Effect of lead on length of root, stem and leaves.

Treatment	Primary root	Stem	Leaves
0 μm	6.13 cm \pm 3.302 100%	14.72 cm \pm 1.23 100%	0.63 cm \pm 0.160 100%
25 μm	5.29 cm \pm 3.045 86.29%	12.90 cm \pm 1.86 12.34%	0.54 cm \pm 0.155 85.71%
50 μm	4.02 cm \pm 2.035 65.6%	12.56 cm \pm 0.87 14.66%	0.54 cm \pm 0.196 85.71%
75 μm	4.24 cm \pm 2.433 69.2%	10.79 cm \pm 1.20 26.65%	0.45 cm \pm 0.133 71.43%
100 μm	4.73 cm \pm 2.776 77.2%	11.75 cm \pm 4.08 20.14%	0.52 cm \pm 0.251 82.54%

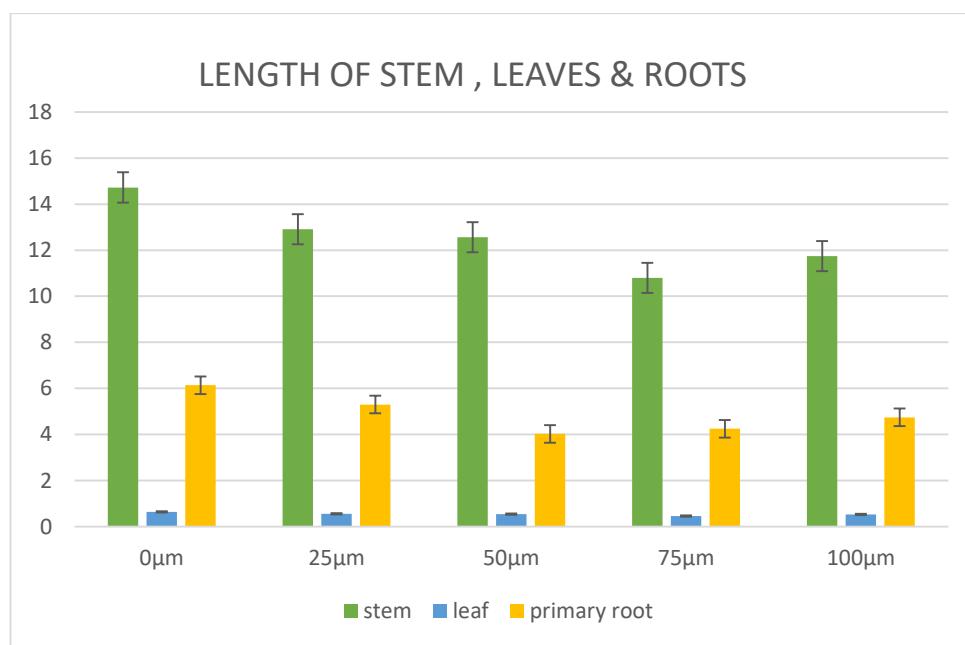


Fig. 1: Effect of lead on the length of root, stem and leaves

Table 2
Lead effect on fresh wt. and dry wt.

Conc.	Fresh weight	Dry weight
0μm	9.00 ± 2.9 100%	1.37 ± 0.10 100%
25 μm	7.81 ± 2.31 86.78%	1.39 ± 0.08 101.46%
50 μm	7.48 ± 2.36 83.11%	1.33 ± 0.05 97.08%
75 μm	7.26 ± 2.24 80.67%	1.33 ± 0.07 97.08%
100 μm	8.82 ± 4.14 98.00%	1.45 ± 0.3 105.84%

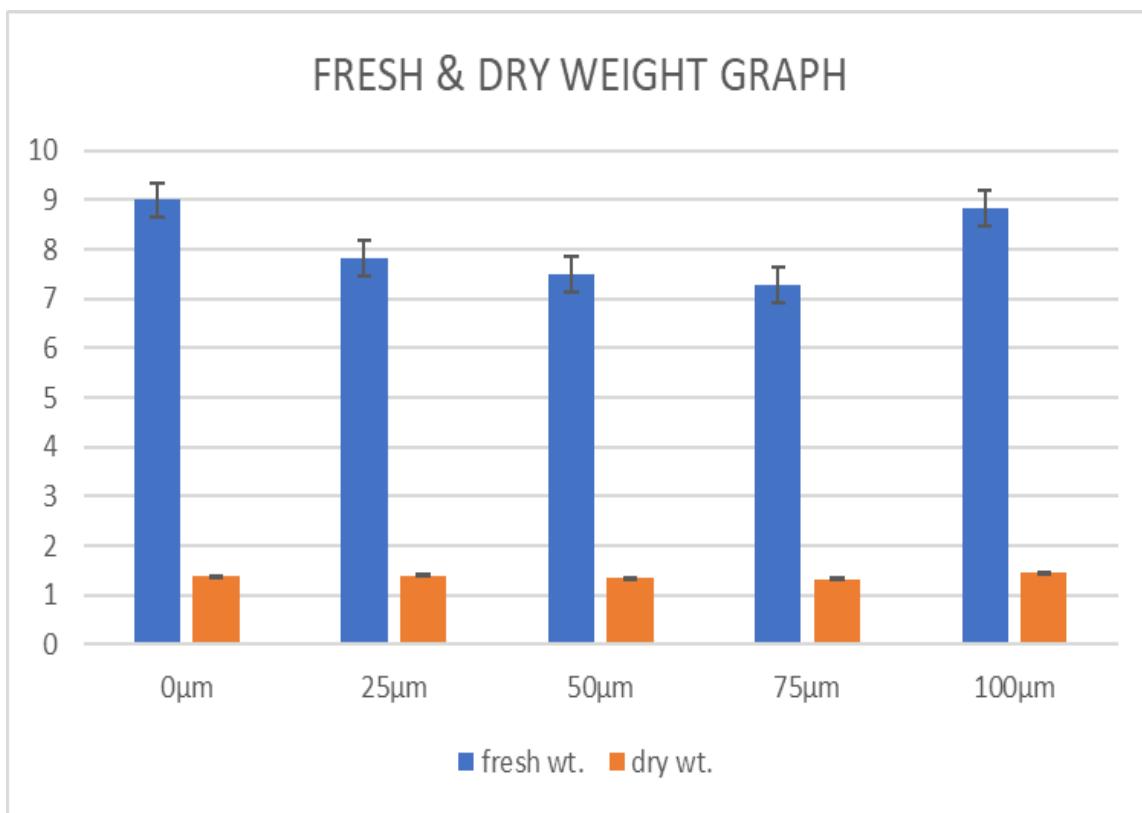


Fig. 2: Effect of lead on fresh wt. and dry wt.

Table 3
Lead effect on chlorophyll content

Treatment	Total chlorophyll	Chlorophyll a	Chlorophyll b
0μm	5.33±6.08 100%	3.47 ± 3.29 100%	1.92 ± 1.61 100%
25μm	2.69±1.42 50.47%	0.67 ± 1.06 19.31%	5.88 ± 6.33 306.25%
50μm	4.01±5.82 75.28%	2.50 ± 3.01 72.05%	1.50 ± 1.73 78.12%
75μm	5.55±7.39 104.12%	3.26 ± 3.85 93.95%	2.33 ± 2.16 121.35%
100μm	2.68±3.49 50.38%	1.48 ± 1.51 42.64%	1.30 ± 1.27 67.71%

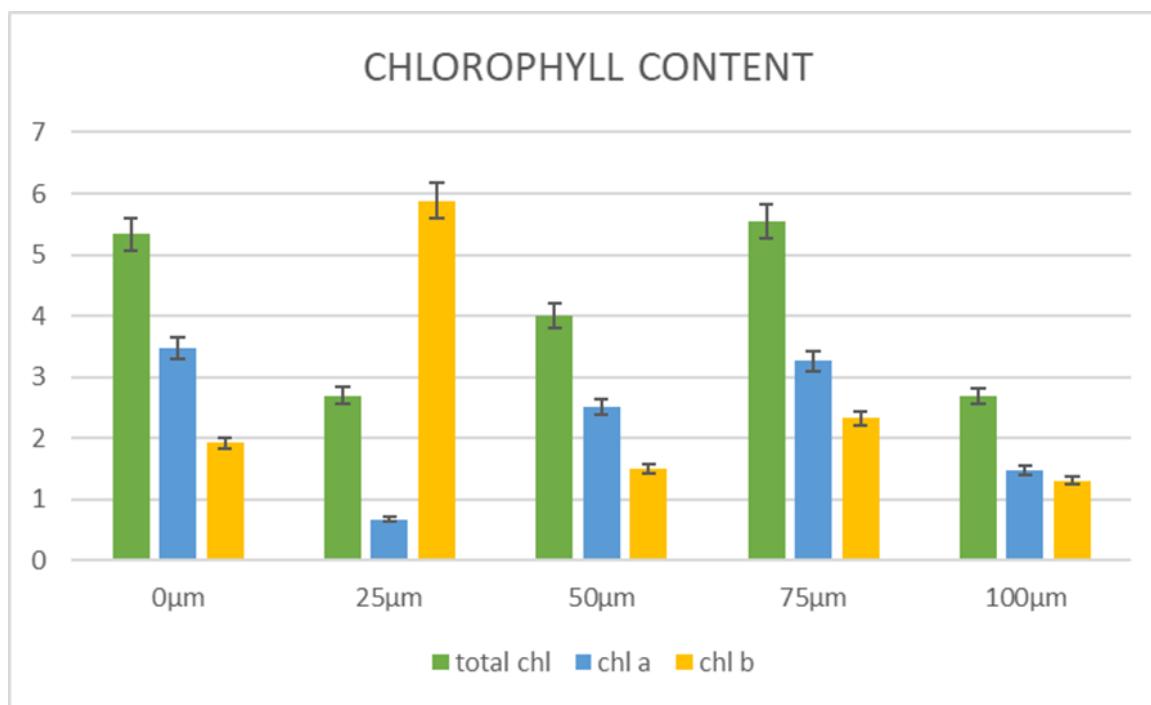


Fig. 3: Effect of lead on chlorophyll content

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(Received 06th January 2025, accepted 08th March 2025)